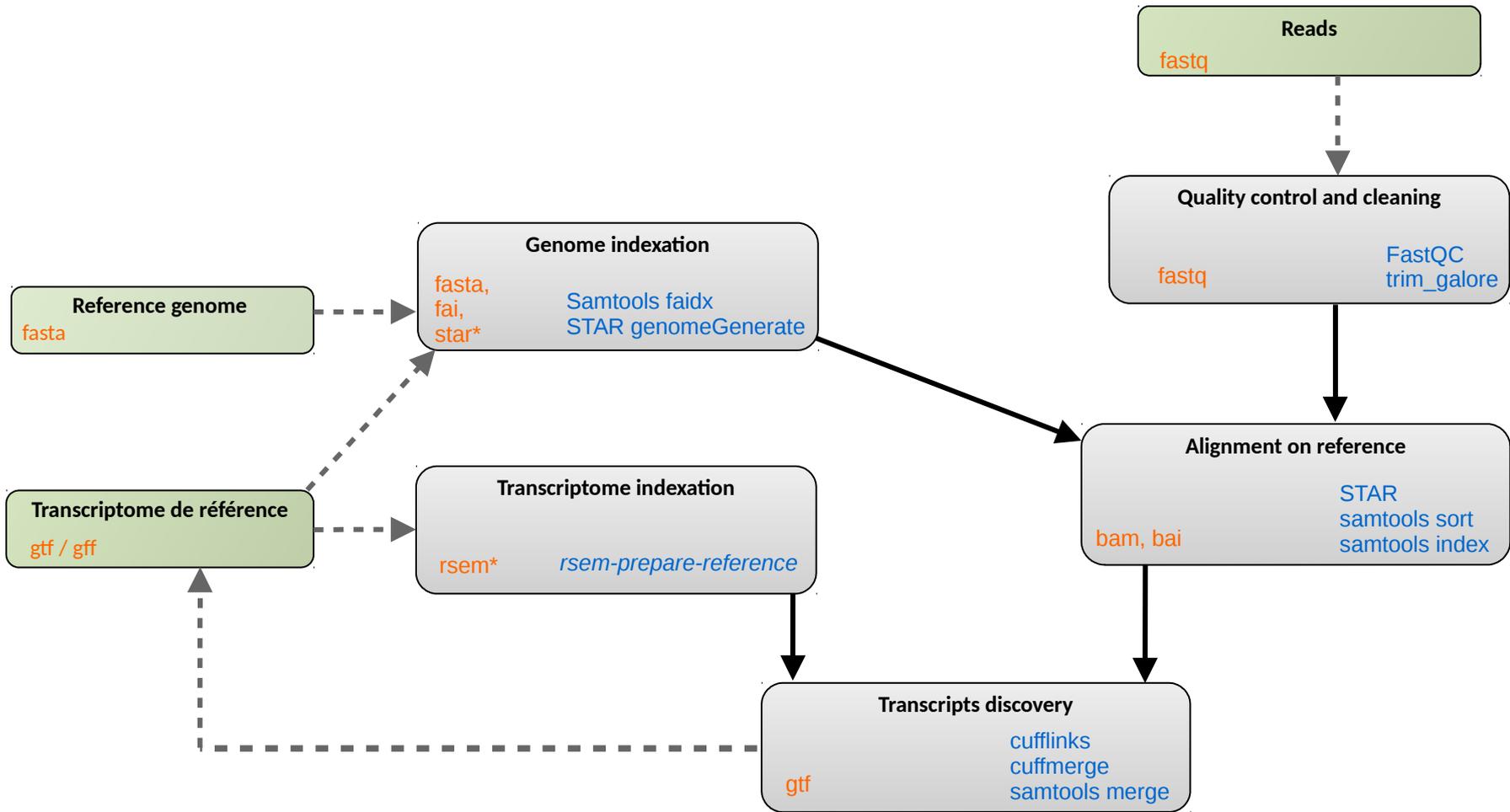


Analysis workflow



Summary - mRNA calling & model comparison

- How to reconstruct transcript ?
- Cufflinks
- Compare models (cuffcompare)
- Merge annotation (cuffmerge)
- Which strategy ?

Transcript reconstruction



Gene location



Exon location



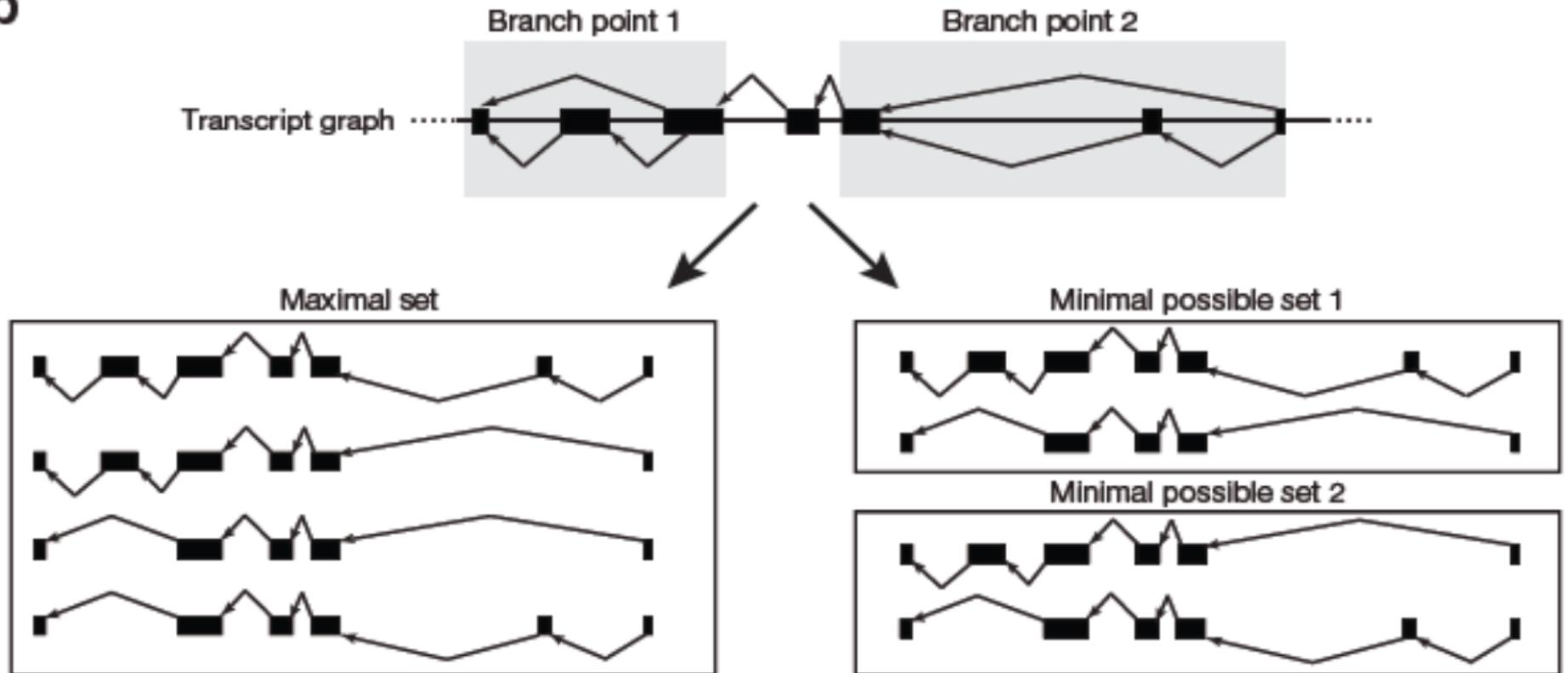
Junctions :

- between read pair junction
- within read junction



Model building strategies

b



REVIEW

Computational methods for transcriptome annotation and quantification using RNA-seq

Manuel Garber¹, Manfred G Grabherr¹, Mitchell Guttman^{1,2} & Cole Trapnell^{1,3}



日本語要約

Transcript assembly and quantification by RNA-Seq reveals unannotated transcripts and isoform switching during cell differentiation

Cole Trapnell, Brian A Williams, Geo Pertea, Ali Mortazavi, Gordon Kwan, Marijke J van Baren, Steven L Salzberg, Barbara J Wold & Lior Pachter

[Affiliations](#) | [Contributions](#) | [Corresponding author](#)

Nature Biotechnology **28**, 511–515 (2010) | doi:10.1038/nbt.1621

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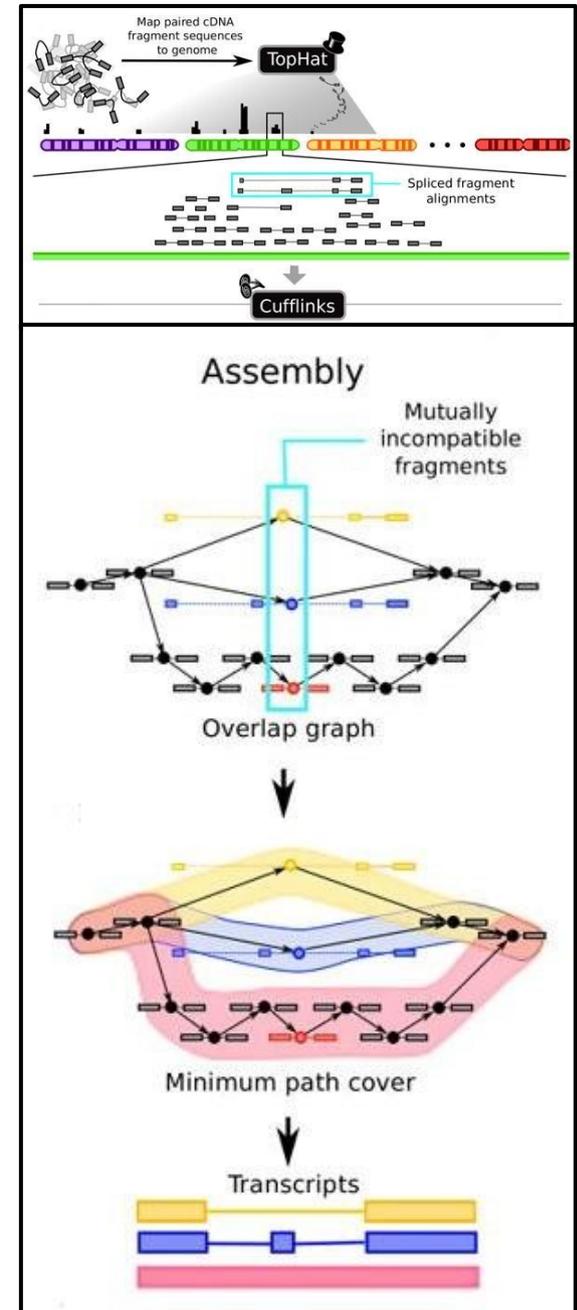
Cufflinks

<http://cole-trapnell-lab.github.io/cufflinks/>

- **assembles transcripts**
- estimates their abundances: based on how many reads support each one
- Suite of software : cufflinks, cuffmerge, cuffcompare

Cufflinks transcript assembly

- Transcripts assembly:
 - fragments are divided into non-overlapping loci
 - each locus is assembled independently
- Cufflinks assembler
 - find the mini nb of transcripts that explain the reads
 - find a minimum path cover (Dilworth's theorem):
 - nb incompatible read = mini nb of transcripts needed
 - each path = set of mutually compatible fragments overlapping each other





Cufflinks inputs and options

```
module load bioinfo/cufflinks-2.2.1
```

- Command line:

```
cufflinks [options] <aligned_reads.(sam/bam)>
```

- Some options:

```
-h/--help
```

```
-o/--output-dir
```

```
-p/--num-threads
```

```
-G/--GTF <reference_annotation.(gtf/gff)>
```

estimate isoform expression, no novel transcripts

```
-g/--GTF-guide <reference_annotation.(gtf/gff)>
```

use reference transcript annotation to guide assembly

```
--max-bundle-length [3,500,000]
```

```
--max-bundle-frags [500,000]
```

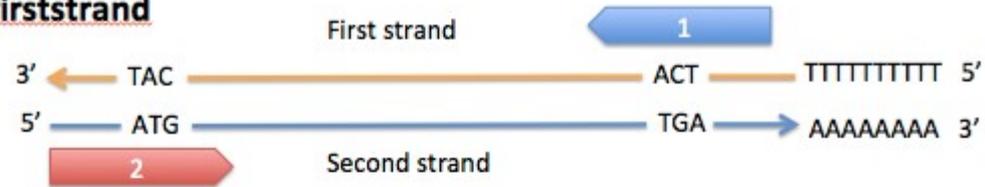
```
--library-type
```

library prep used for input reads

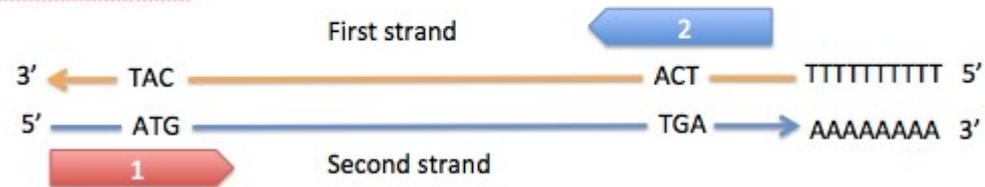


Cufflinks library types

fr-firststrand



fr-secondstrand



Mix of both = fr-unstranded



Cufflinks outputs

- `transcripts.gtf`
contains assembled isoforms (coordinates and abundances)
- `genes.fpkm_tracking`
contains the genes FPKM
- `isoforms.fpkm_tracking`
contains the isoforms FPKM
- `skipped.gtf`
contains skipped loci (too many fragments)



Cufflinks GTF description

`transcripts.gtf` (coordinates and abundances):

- contains assembled isoforms
 - can be visualized with a genome viewer
 - attributes: ids, FPKM, confidence interval, read coverage & support
- score: most abundant isoform = 1000
minor isoforms = minor FPKM/major FPKM
 - cov: estimate for depth across the transcript

```
1 Cufflinks transcript 459812 460830 1 - .
1 Cufflinks exon 459812 460830 1 - .
1 Cufflinks transcript 463572 478996 1000 - .
1 Cufflinks exon 463572 463746 1000 - .
1 Cufflinks exon 466228 466405 1000 - .
```

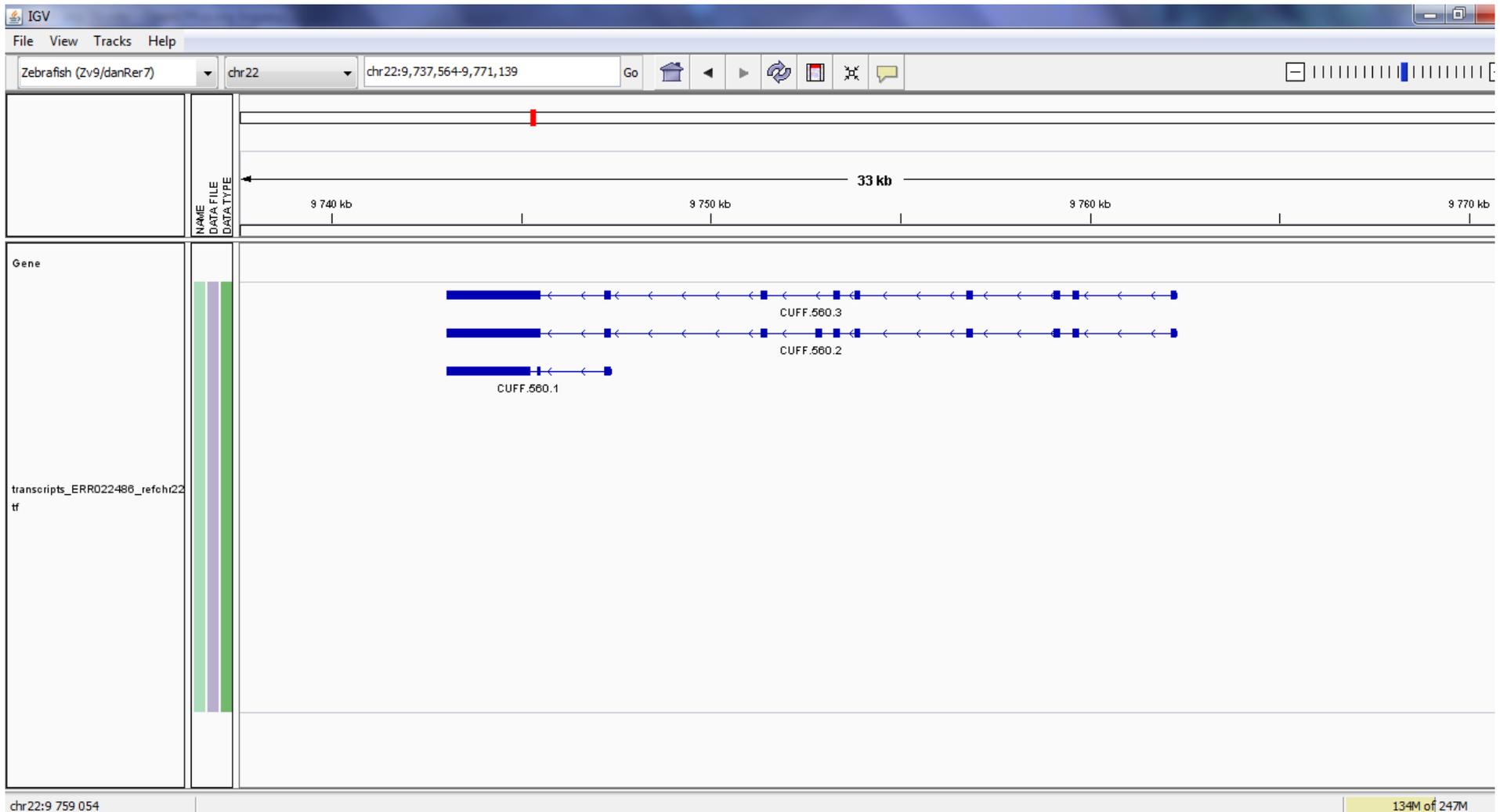
```
gene_id "ENSBTAG00000013841"; transcript_id "ENSBTAT00000018387"; FPKM "0.0000000000"; frac "0.000000";
gene_id "ENSBTAG00000013841"; transcript_id "ENSBTAT00000018387"; exon_number "1"; FPKM "0.0000000000"; frac "0.000000";
gene_id "CUFF.2"; transcript_id "ENSBTAT00000015319"; FPKM "25.4745974237"; frac "1.000000";
gene_id "CUFF.2"; transcript_id "ENSBTAT00000015319"; exon_number "1"; FPKM "25.4745974237"; frac "1.000000";
gene_id "CUFF.2"; transcript_id "ENSBTAT00000015319"; exon_number "2"; FPKM "25.4745974237"; frac "1.000000";
```

```
conf_lo "0.000000"; conf_hi "0.000000"; cov "0.000000"; full_read_support "no";
conf_lo "0.000000"; conf_hi "0.000000"; cov "0.000000";
conf_lo "21.387219"; conf_hi "29.561976"; cov "422.904985"; full_read_support "yes";
conf_lo "21.387219"; conf_hi "29.561976"; cov "422.904985";
conf_lo "21.387219"; conf_hi "29.561976"; cov "422.904985";
```



Cufflinks GTF description

transcripts.gtf (coordinates and abundances):
visualization in IGV





Cufflinks / Cuffcompare

Compare assemblies between conditions:

- compare your assembled transcripts to a reference annotation
- track Cufflinks transcripts across multiple experiments

Command:

```
cuffcompare [-r <reference.gtf>] [-o <outprefix>] <input1.gtf>  
...
```

Outputs:

- `<outprefix>.stats` - overall summary statistics
- `<outprefix>.combined.gtf` - “union” of all transfrags
- `<cuff_in>.refmap` - transfrags matching to reference transcript
- `<cuff_in>.tmap` - best reference transcript for each transfrag
- `<outprefix>.tracking` - tracking transfrags across samples

Class code de cuffcompare

=	complete match	
c	contained	
j	novel isoform	
e	single exon	
i	within intron	
o	exonic overlap	
p	polymerase run-on	
r	repeat	
u	unknown, intergenic	
x	exonic overlap on the opposite strand	
s	intronic overlap on the opposite strand	



Cufflinks / Cuffmerge

Merge together several assemblies:

- merge novel isoforms and known isoforms
- filters a number of transfrags that are probably artifacts
- build a new gene model describing all conditions

Command:

```
cuffmerge [options] -o <assembly_GTF_list>
```

Options:

- `-o/ --output-dir`
- `-g/ --ref-gtf`
- `-s/ --ref-sequence`
- `--min-isoform-fraction`
discard isoforms with abundance below this [0.05]
- `-p/ --num-threads`



Cufflinks / Cuffmerge

merged.gtf (coordinates and legacy):

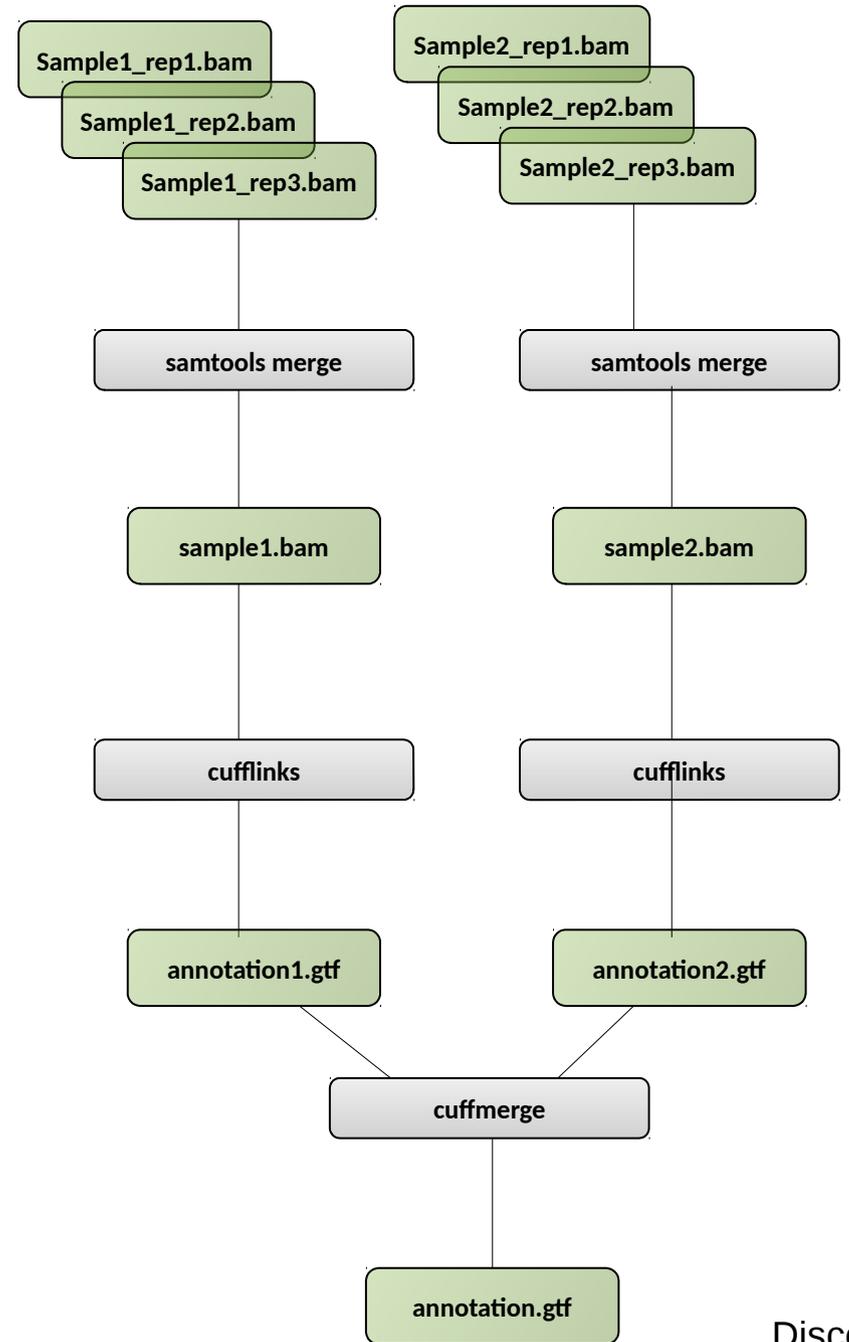
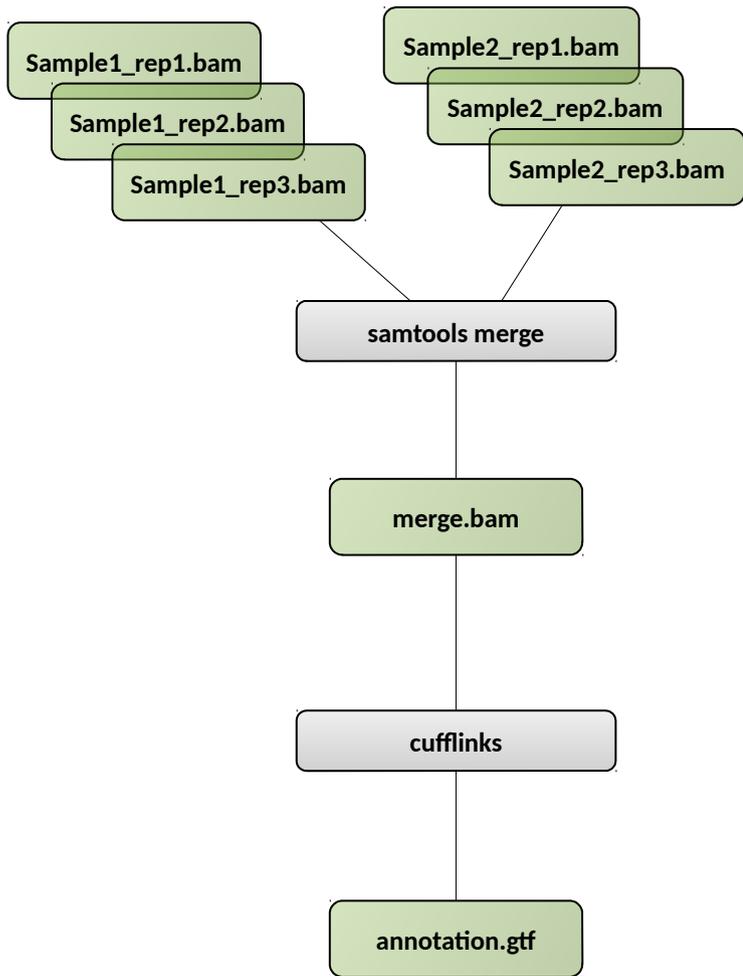
- contains merged input assemblies
- can be visualized with a genome viewer
- attributes: ids, name, old, nearest_ref, class_code, tss_id, p_id

```
1 Cufflinks exon 34627 35558 . + .
1 Cufflinks exon 242394 242646 . + .
1 Cufflinks exon 275623 275681 . + .
1 Cufflinks exon 242402 242646 . + .
1 Cufflinks exon 254559 254693 . + .
1 Cufflinks exon 247340 249673 . + .
1 Cufflinks exon 351546 351874 . + .
1 Cufflinks exon 355064 355237 . + .
1 Cufflinks exon 357793 357952 . + .
1 Cufflinks exon 361144 362915 . + .
```

```
gene_id "XLOC_000001"; transcript_id "TCONS_00000001"; exon_number "1"; gene_name "ENSBTAG00000006858";
gene_id "XLOC_000002"; transcript_id "TCONS_00000002"; exon_number "1"; gene_name "CBX3";
gene_id "XLOC_000002"; transcript_id "TCONS_00000002"; exon_number "2"; gene_name "CBX3";
gene_id "XLOC_000002"; transcript_id "TCONS_00000003"; exon_number "1";
gene_id "XLOC_000002"; transcript_id "TCONS_00000003"; exon_number "2";
gene_id "XLOC_000003"; transcript_id "TCONS_00000004"; exon_number "1";
gene_id "XLOC_000004"; transcript_id "TCONS_00000005"; exon_number "1"; gene_name "RCAN1";
gene_id "XLOC_000004"; transcript_id "TCONS_00000005"; exon_number "2"; gene_name "RCAN1";
gene_id "XLOC_000004"; transcript_id "TCONS_00000005"; exon_number "3"; gene_name "RCAN1";
gene_id "XLOC_000004"; transcript_id "TCONS_00000005"; exon_number "4"; gene_name "RCAN1";
```

```
oId "ENSBTAT00000009004"; nearest_ref "ENSBTAT00000009004"; class_code "="; tss_id "TSS1";
oId "CUFF.1.1"; nearest_ref "ENSBTAT00000007283"; class_code "x"; tss_id "TSS2";
oId "CUFF.1.1"; nearest_ref "ENSBTAT00000007283"; class_code "x"; tss_id "TSS2";
oId "CUFF.1.2"; class_code "u"; tss_id "TSS2";
oId "CUFF.1.2"; class_code "u"; tss_id "TSS2";
oId "CUFF.2.1"; class_code "u"; tss_id "TSS3";
oId "CUFF.3.1"; nearest_ref "ENSBTAT00000037243"; class_code "j"; tss_id "TSS4";
```

Which strategy ?





Hands-on: transcripts assembly

Using cufflinks:

Exercise 6:

- reconstruct known and novel transcripts
- compare annotations